





A anatomia como ferramenta para a identificação da casca de Pterocarpus angolensis e

Terminalia sericea

Teresa Quilhó¹; Fernanda Bessa¹; Ana Isabel Ribeiro-Barros²; Natasha Ribeiro³

¹ Centro de Estudos Florestais (CEF)/ Instituto Superior de Agronomia/Universidade de Lisboa; <u>terisantos@isa.ulisboa.pt</u>;² Centro de Investigação em Agronomia, Alimentos, Ambiente e Paisagem (LEAF)/ Instituto Superior de Agronomia/Universidade de Lisboa; ³ Faculdade de Agronomia e Engenharia Florestal/ Universidade Eduardo Mondlane

RESUMO: *Pterocarpus angolensis* e *Terminalia sericea* são espécies africanas com potencial medicinal. Apesar da importância da sua casca como um adstringente poderoso para tratar várias doenças, a sua descrição é escassa. Para a correta identificação e padronização do material vegetal, foram coletadas amostras de casca de cada espécie e analisadas ao microscópio ótico e eletrônico. Consideraram-se como características importantes para a sua identificação: o esclerênquima, maioritariamente sob a forma de fibroesclercídos em *P. angolensis* e as células secretoras de grande dimensão dispostas em linhas cu faixas tangenciais distintas logo no mício do floema condutor; células cristaliferas (drusas) em linhas tangenciais regulares e ocorrência de um enorme cristal, junto ou incluso nas faixas tangenciais de fibras em *T. sericea*. Os resultados obtidos provam que a anatomia da casca pode ser utilizada como um importante subsídio na identificação e estandardização das espécies podendo o conhecimento científico contribuir para formas de fiscalização mais efetivas na prevenção da adulteração das espécies no comércio.

Palavras-chaves: Floema secundário, Estrutura anatómica, Distinção de espécies, Plantas medicinais.

The anatomy as a tool for the identification of the bark of *Pterocarpus angolensis* and

Terminalia sericea

ABSTRACT: *Pterocarpus angolensis* and *Terminalia sericea* are two African species with medicinal potential. Despite the importance of their bark as a powerful astringent to treat various diseases it is poor described. In order to provide referential information for correct identification and standardization of the plant material, bark samples from each two species were collected and analyzed under light and electron microscopy. Some important anatomical features to identification were: the sclerenchyma tissue mostly in form of fibre-sclereids and the large secretory cells arranged in conspicuous rows or tangential bands in the conducting phloem in *P. angolensis;* the



KAAPIR











crystalliferous cells arranged in very regular tangential rows (druses) and the occurrence of large crystal cells near or including the tangential fibre bundles in *T. sericea* bark.

The results obtained show that the anatomy of the bark can be used as an important subsidy in identification and standardization of the studied species contributing the scientific knowledge for more effective forms of scrutiny in preventing commercial adulteration of species.

Keywords: Secondary phloem, Anatomical structure, Species distinction, Medicinal plants.

1. INTRODUCTION

Bark has a major role as a sources of natural products used in traditional medicine. In recent years experimental studies on bark show their potential bioactivity (Carmo et al., 2016). In order to contribute to the plant identification the present study reports a brief anatomical qualitative bark description of *Pterocarpus angolensis* DC. and *Terminalia sericea* Burch. ex DC., two popular medicinal species in the miombo woodlands (Moura et al.2017, Mongalo et al., 2016).

P. angolensis is a deciduous tree belonging to the family Fabaceae. The bark is used as a powerful astringent to treat the diarrhoea, nose bleeding, stomach-disorders, gonorrhea, and skin lesions (Moura et al. 2017). *T. sericea* is a deciduous tree belonging to the family of Combretaceae; pulverized bark is applied to wounds and taken to treat diabetes, cough, sore throat and stomach-ache (Lemmens, 2009). Adulteration of botanical material in commerce is common and microscopic evaluation of bark via light microscopy and scanning electron microscopy (SEM) provides referential information for correct identification and standardization of the plant material (Serrano et al., 2010). Anatomical descriptions are the first step towards establishing the identity of raw material, but the knowledge on bark characteristics is limited to a small number of species. There is any reference for *P. angolensis* and *T. sericea*. Recently studies on *Terminalia* sp. e.g. *T. arjuna* (Sivaji et al., 2012), *T. chebula* (Ingle & Dhabe, 2015) enlightening the bark diagnostic features.

The objective of this study was to analyze the anatomy of bark of *P. angolensis* DC and *T. sericea* Burch. ex DC. for its correct identification. The findings from this study would be useful as standards for the species as well as a source of reference for further scientific investigation on the species enabling the diffusion of scientific knowledge to the society.

2. MATERIAL AND METHODS

The anatomical studies were conducted on the barks samples of P. angolensis, and T. sericea













growing in northern Mozambique (Miombo). Bark samples were collected at Dbh (1.30 m above ground) using a cutlass and then removing it from the trunk by tapping with a hammer. In the laboratory samples were impregnated with DP1500 polyethylene glycol according Barbosa et al. (2010). Transverse and longitudinal microscopic sections of approximately 17 μ m thicknesses were prepared with a Leica SM 2400 microtome. The sections were stained with a double staining of chrysodine/astra blue and were mounted on Kaiser glycerin. After 24 h drying, the lamellas were submerged into xylol for 30 minutes, dehydrated on 96% and 100% alcohol and mounted on Eukitt. Individual specimens were also taken sequentially from the cambium towards the periphery and were macerated in a 1:1 solution of 30% H₂O₂ (hydrogen peroxide) and CH₃COOH (glacial acetic acid) at 60°C for 48 h and stained with astra blue. Light microscopic observations were made using Leica DM LA and photomicrographs were taken with a Nikon Microphot-FXA. Small samples with approximately 5 mm of edge were cut with a sharp razor blade, and their surfaces were examined with a scanning electron microscope (SEM) Hitachi TM 3030 Plus at 5 kV with different magnifications, and the images were recorded in digital format. The bark terminology followed Angyalossy et al. (2016).

3. RESULTS

Pterocarpus angolensis DC

Secondary phoem is non-layered and the secretory cells produce the main pattern. The secretory cells are large distributed within the entire secondary phoem arranged in conspicuous rows or tangential bands (Figure 1A, 2A-C) appearing early in the conducting phoem. The secretory cells are filled with a prominent muss of reddish content. The sieve tubes elements (conducting cells) are mostly isolated and appeared interspersed within the axial parenchyma (storage tissue) which formed bands enclosing the tangential bands of secretory cells and sclerenchyma tissue (mechanical tissue) throughout the phloem. The sclerenchyma tissue are mostly in form of sclereids and fibre-sclereids arranged in short or long tangential clusters, mixed with few fibres. The sclereids had different shape and size, round to rectangular and the fibre-sclereids had short cell body and lateral projections with blunt or pointed end walls, both with thick walls (Figure 2B-C). Rays are uniseriate, homogeneous with procumbent cells and stratified. Rays (storage/conducting tissue) distorted slightly near the sclerenchyma and/or the secretory cell bundles or enlarged shortly within the nonconducting phloem toward outside. Crystals of calcium oxalate appeared mostly in forms of prisms occurring in series of axial parenchyma (Figure 2C).

PATROCINADORES:







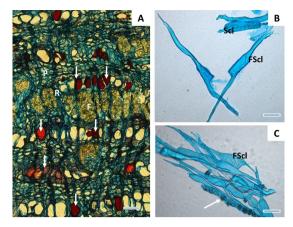












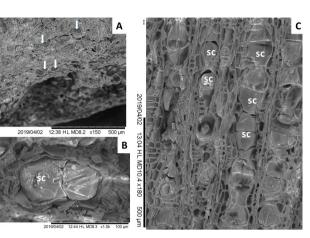


Figure. 1- Secondary phloem of *P. angolensis*. A) Transverse section; B) individualized cells. Rays (R); fibers (F); axial parenchyma (P); secretory cells (arrows in A) and crystals (arrows in C); sclereids (Scl); fibre-sclereids (Fscl). Scale bar= $100 \ \mu m$

Figure 2 - Secondary phloem of *P. angolensis* (SEM). A) Transverse section; secretory cells (arrows); B) large secretory cell (Sc); C) secretory cells (Sc), radial section.

Terminalia sericea Burch. ex DC

Secondary phloem is layered and almost regular from cambium to the periphery. The conducting phloem includes layers of tangential bands of parenchyma cells and sieve tube elements alternating with groups or tangential bands of fibres. The axial parenchyma cells are arranged in various seriate tangential bands: rows of crystalliferous parenchyma cells arranged in very regular tangential rows are followed by rows of normal parenchyma cells as seen in transverse section (Figure 3A). On the transverse section the fibres are arranged regularly in short alternating plates mixed with long tangential bands somewhat undulated. The rays are non-storied, mostly uniseriate sometimes biseriate of procumbent cells; rays are thin when cross the fibres bundles but between them rays start to dilate by inflation or cell division. Sclereids and expanded axial parenchyma cells arise from parenchyma cells and are observed near/or including the fibre bundles (Figures. 3A-C).

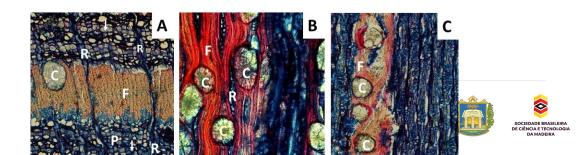








Figure 3. Secondary phloem of *T. sericea*. A) Transverse section; B) tangential section; C) radial section; rays (R); fibers (F); axial parenchyma (P); sieve tubes (S); large crystal (C) and crystals in axial parenchyma (arrows). Scale bar=100 μ m

Also druses in chambered parenchyma cells are arranged in very regular tangential rows as seen in transverse section (Figure. 3A and Figure, 4A-C). Organic content, presumably phenolic compounds are observed in parenchyma tissue.

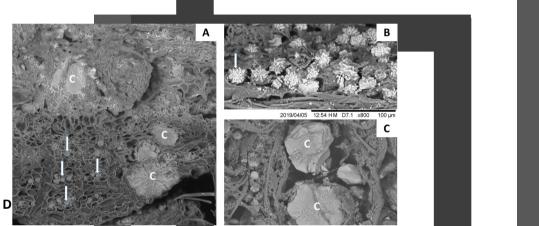


Figure 4 Secondary phloem of *T. sericea* (SEM). A) Crystals in parenchyma cells in tangential rows (arrows, druses), B) druses (arrows) and C) large crystals (c).

4. DISCUSSION

Bark comprises all tissues outside the vascular cambium including the secondary phloem, the periderm and rhytidome (Angyalossy et al., 2016), therefor has a highly complex and heterogeneous structure. According to our results, the general anatomy of *P. angolensis* (Figure 1,2) *and T. sericea* (Figure 3,4) bark had some similarities with other species of *Pterocarpus, Terminalia* and other genus of the family Fabaceae and Combretaceae detailed described by Roth (1981). In our study particular attention was given on the development of the secretory system and the form, shape and occurrence of crystal cells in the secondary phloem. According Alamgir (2017) the powdered crude drugs can be identified based on the form, the presence or absence of different cell types and cell inclusions. In this sense recent studies were conducted with different species helpful

















for its discrimination ensuring safety for commercial pharmacological uses e.g. *Acacia suma* Roxb (Dash et al. 2014) and *Erythrina* × *neillii* Mabberley & Lorence (Gabr et al. 2017).Both species were distinguished by the secretory structures and type of sclerenchyma in *P. angolensis* and by crystal cells in *T. sericea*. Secretory cells were reported in Fabaceae e.g. *Centrolobium paraense* Tul. or in *Machaerium* sp.(Roth, 1981) and the regular arrangement of crystal cells imposing a conspicuous pattern in secondary phloem of Combretaceae was also stated by various authors .e.g. in *T. arjuna* (Sivaji et al., 2012).

The present study shows the significance of anatomical analysis as a tool for an accurate identification of the two species and may serve as a reference point for other new researches in order to disseminate the scientific knowledge of plants widely used in traditional medicine.

5. CONCLUSION

The anatomical structure of secondary phloem of *Pterocarpus angolensis* and *Terminalia sericea* was analyzed for the first time. The barks of the two species were markedly different and distinguished by the type of sclerenchyma and the large secretory cells arranged in conspicuous rows or tangential bands in *P. angolensis* and by the arrangement of the crystalliferous cells and occurrence of large crystal cells near or including the fibre bundles and in *T. sericea*.

The present study proves the significance of anatomical analysis as a tool for identification of these plants. It will be useful for the authentication of herbal products avoiding the presence of adulterants and thereby contributing to the scientific world of research.

6. REFERENCES

Alamgir A.N.M. Therapeutic use of medicinal plants and their extracts. Vol. 1 Pharmacognosys. Springer; 2017

Angyalossy V, Pace MR, Evert RF, Marcati CR, Oskolski AA, Terrazas T. et al.. IAWA List of microscopic bark features. International Association of Wood Anatomists Journal 2016; 37: 517–615.

Barbosa ACF, Pace MR, Witovisk L, Angyalossy V. A new method to obtain good anatomical slides of heterogenerous plant parts. International Association of Wood Anatomists Journal 2010; 31: 373-383.

Carmo JF, Miranda I, Quilhó T, Sousa VB, Cardoso S, Carvalho AM. Chemical and structural characterization of the bark of Albizia niopoides trees from the Amazon. Wood Science Technology 2016; 50 (4): 677–692.

Dash G.K., Abdullah M.S, Acharyya S. Pharmacognostic Evaluation of the Bark of Acacia suma Roxb (Fabaceae).Journal of Pharmaceutical Research 2014; 13 (6): 961-966



















Gaber S.K., Bakr R.O., Elshishtawyb H.M., El-Fishawyc A.M., El-Alfy T.S. Botanical and genetic characters of Erythrina × neillii cultivated in Egypt. Revista Brasileira de Farmacognosia. 2017; 27:273-281

Ingle P, Dhabe A. Anatomical investigation of Terminalia chebula Retz. Phytotaxonomy. 2015; Vol. 15: 55-62.

Lemmens RHM. Terminalia sericea Burch. ex DC. In: Lemmens RHMJ, Louppe D, Oteng-Amoako AA, editor. PROTA (Plant Resources of Tropical Africa / Ressources végétales de l'Afrique tropicale). Wageningen, Netherlands; 2009.

Mongalo N.I., McGaw L.J., Segapelo T.V., Finnie J.F., Staden J.Van. Ethnobotany, phytochemistry, toxicology and pharmacological properties of Terminalia sericea Burch. ex DC. (Combretaceae) -A review. Journal of Ethnopharmacology. 2016; 194: 789-802

Moura I, Maquia I, Rija AA, Ribeiro N, Ribeiro-Barros AI. Biodiversity studies in key species from the African Mopane and Miombo Woodlands. In: Bitz L, ed. Genetic Diversity. Rijeka: InTech; 2017.

Roth I. Structural patterns of tropical barks. Encyclopedia of plant anatomy Vol. LX, Part 3. Gebruder Borntraeger, Berlin; 1981

Serrano R, Silva G, Silva O. Application of Light and Scanning Electron Microscopy in the Identification of Herbal Medicines. In: A Méndez-Vilas, J. Díaz, editor. Microscopy: Science, Technology, Applications and Education. Badajoz Spain; 2010.

Sivaji K, Mahendra M, Ramesh L, Madhava K. Comparative pharmacognostical studies of Terminalia arjuna used in ayurvedic drug "arjuna" with its adulterant Kavalama urens. Indian Journal of Plant Sciences 2012; Vol. 1 (2&3): 229-238.

Takawira-Nyenya R. Pterocarpus angolensis. In: Jansen PCM, Cardon D, editor. Plant Resources of Tropical Africa 3. Dyes & Tannins. PROTA Foundfation, Wageningen, Netherlands/Backhugs Publishers, Leiden, Netherlanmds/CTA, Wageningen, Netherlands; 2005













